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3 **Biological Cybernetics manuscript No.**  
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10 Michele Giugliano · Giancarlo La Camera · Stefano Fusi · Walter Senn

# 11 **The response of cortical neurons to *in vivo*-like input current: theory** 12 **and experiment**

## 13 **II. Time-varying and spatially distributed inputs**

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23 **Abstract** The response of a population of neurons to time-  
24 varying synaptic inputs can show a rich phenomenology,  
25 hardly predictable from the dynamical properties of the mem-  
26 brane's inherent time constants. For example, a network of  
27 neurons in a state of spontaneous activity can respond sig-  
28 nificantly more rapidly than each single neuron taken in-  
29 dividually. Under the assumption that the statistics of the  
30 synaptic input is the same for a population of similarly be-  
31 having neurons (mean field approximation), it is possible  
32 to greatly simplify the study of neural circuits, both in the  
33 case in which the statistics of the input are stationary (re-  
34 viewed in La Camera et al 2008) and in the case in which  
35 they are time-varying and unevenly distributed over the den-  
36 dritic tree. Here we review theoretical and experimental re-  
37 sults on the single-neuron properties that are relevant for  
38 the dynamical collective behavior of a population of neu-  
39 rons. We focus on the response of integrate-and-fire neurons  
40 and real cortical neurons to long-lasting, noisy, *in vivo*-like  
41 stationary inputs and show how the theory can predict the

observed rhythmic activity of cultures of neurons. We then  
show how cortical neurons adapt on multiple time-scales in  
response to input with stationary statistics *in vitro*. Next,  
we review how it is possible to study the general response  
properties of a neural circuit to time-varying inputs by es-  
timating the response of single neurons to noisy sinusoidal  
currents. Finally, we address the dendrite-soma interactions  
in cortical neurons leading to gain modulation and spike  
bursts, and show how these effects can be captured by a two-  
compartment integrate-and-fire neuron. Most of the experi-  
mental results reviewed in this article have been successfully  
reproduced by simple integrate-and-fire model neurons.

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### 1 Introduction

Neurons in the intact brain of the mammalian cortex are  
driven by the synaptic current generated by thousands of  
other neurons. As their activity is determined by the spikes  
of large populations of neurons, it is important to study and  
characterize the collective properties of groups of cells. When  
similarly behaving neurons share approximately the same  
statistics of the synaptic current, these neurons can be grouped  
together, and they can be replaced by a single representative  
cell. This approach, known as the “mean field approxima-  
tion”, is reviewed in La Camera et al (2008) for the case in  
which i) the statistics of the input current is stationary and ii)  
the response of the neuron is quasi-static, i.e. it can be con-  
sidered constant on a time interval of a few seconds. This  
approach can be extended to the more general case of time-  
varying statistics of the synaptic currents, and to the case  
where the output spike train is non-static. The extended ap-  
proach is particularly important to describe transient, oscil-  
latory and rhythmic neuronal activity of large populations of  
neurons, and it can predict dynamical behaviors that some-  
times cannot be foreseen by looking at the passive properties  
of the neuronal membrane. Population response times can be  
very different from those predicted by the inherent time con-  
stants of every individual neuron and in general they depend  
on the full distribution of all dynamical variables character-  
izing the neuronal dynamics. A population of neurons with

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M. Giugliano acknowledges support from the EPFL and the Euro-  
pean Commission (FACETS and EUSynapse Project); G. La Camera  
is supported by the Intramural Research Program, NIMH. S. Fusi is  
supported by the Swiss National Science Foundation, grant PP00A-  
106556. W. Senn is supported by the Swiss National Science Founda-  
tion, grant No 3152A0-105966. The views expressed in this article do  
not necessarily represent the views of the NIMH, NIH, or the United  
States Government.

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M. Giugliano  
Laboratory of Neural Microcircuitry, Ecole Polytechnique Fédérale de  
Lausanne, Station 15, CH-1015 Lausanne, Switzerland, and Theoret-  
ical Neurobiology, University of Antwerp, Wilrijk, Belgium. Tel.: +41-  
21-6931658, Fax: +41-21-6935350.  
E-mail: michele@tmb.ua.ac.be

G. La Camera  
Laboratory of Neuropsychology, National Institute of Mental Health,  
National Institutes of Health, Bethesda, USA.

S. Fusi  
Center for Theoretical Neuroscience, Columbia University School of  
Medicine, New York, USA, and Institute of Neuroinformatics, Univer-  
sity of Zurich - ETH, Zurich, Switzerland.

W. Senn  
Department of Physiology, University of Bern, Switzerland.

spontaneous activity will exhibit reaction times that are significantly shorter than the membrane time constant (see, e.g., van Rossum et al (2002)), because the depolarizations are distributed in the whole interval between the resting potential and the threshold for spike emission. As a consequence, some of the neurons are ready to generate an action potential quickly (order of one millisecond) in response to a stimulus. Other single-neuron properties, like adaptation on time-scales of hundreds of milliseconds to seconds, are also readily reflected by the population response. The extension of mean field approach to include non-stationary properties are the focus of this review.

We start by presenting an example where the quasi-stationary response properties of single neurons allow one to make quantitative predictions on the rhythmic activity exhibited by networks of dissociated neurons (Giugliano et al 2004). We then show how single neurons and populations of neurons exhibit firing rate adaptation on multiple time-scales (La Camera et al 2006), even in response to inputs with stationary statistics. Studies of the population response to fast-varying inputs are reviewed next. In such a case, the full mathematical machinery of the so-called ‘‘population density approach’’, based on the solution of a Fokker-Plank equation, would in general be needed (Knight 1972; Abbott and van Vreeswijk 1993; Treves 1993; Fusi and Mattia 1999; Brunel and Hakim 1999; Nykamp and Tranchina 2000). However, recent studies on simple integrate-and-fire (IF) model neurons revealed that it is possible to predict the population response to an arbitrary time-varying input by knowing the response of single neurons to the sinusoidal components that make the input signal (Brunel et al 2001; Fourcaud and Brunel 2002). The single-neuron response can be estimated in the case of IF model neurons, or it can be measured directly for real cortical cells (Köndgen et al 2008). Because of the features of sodium-mediated action potentials, cortical neurons are surprisingly good at relaying fast temporal information, while dynamical response attenuation and distortions affect the input-output transfer properties only for very fast ( $\lesssim 5$  ms) input transients. In all these studies, IF neurons with firing rate adaptation and other small modifications could predict quantitatively many aspects of the behavior of real cortical neurons.

In the last section we turn back to the static properties of the neuronal response function to show how it is modulated by the spatial input distributions along the dendritic tree. We review the experimental results that show how distal dendritic input can modulate in a non-linear fashion the somatic stationary response function (Larkum et al 2004). This modulation is due to an interaction between somatic action potentials and dendritic calcium spikes, that under the right conditions can lead to a further burst of somatic action potentials (Larkum et al 1999). Despite the complexity of these mechanisms, a simple somatic IF mechanism coupled with a dendritic compartment can capture the experimental phenomenology and explain the gain modulation induced by dendritic inputs. The extent to which simple spiking models can capture these complex phenomena and, thus, provide

the field of Neuroscience with a successful reductionist approach is further examined in the Discussion.

### 1.1 Response function of the leaky IF neuron

Most of the material reviewed in this article is based on, or is related to, the response function of the adaptive leaky IF (LIF) neuron, reviewed in detail in the companion article (La Camera et al 2008), to which the reviewer is referred for a more detailed exposition. We report here the main formulae for convenience. The LIF neuron is completely characterized by its membrane potential,  $V$ , which evolves according to

$$\frac{dV}{dt} = -\frac{V - V_{rest}}{\tau} + \frac{I}{C}, \quad (1)$$

until a threshold  $\theta$  is reached. At this time, a spike is said to be emitted and  $V$  is clamped to a reset potential  $V_r$  for a refractory period  $\tau_r$ , after which motion resumes according to Eq. 1.  $V_{rest}$  is the membrane’s resting potential,  $C$  is the membrane capacitance,  $\tau = RC$ , where  $R$  is the membrane resistance, and  $I$  is an input current. To emulate the noisy input current targeting neurons *in vivo*, the current  $I$  is modeled as a stochastic process. In the diffusion approximation (Lánský and Sato 1999; Richardson and Gerstner 2005), the subthreshold dynamics of the membrane potential obeys the stochastic differential equation of the Ornstein-Uhlenbeck process (Tuckwell 1988),

$$dV = -\frac{V - V_{rest}}{\tau} dt + \mu dt + \sigma \xi_t \sqrt{dt}, \quad (2)$$

where

$$\mu = m_I/C, \quad \sigma = \sqrt{2\tau} s_I/C \quad (3)$$

are the average and standard deviation in unit time of the membrane voltage, and  $\xi_t$  is a Gaussian process with flat spectrum and unitary variance.  $m_I$  and  $s_I^2$  are the average and the variance of the synaptic input current, and  $\sqrt{2\tau}$  is a factor to preserve units.

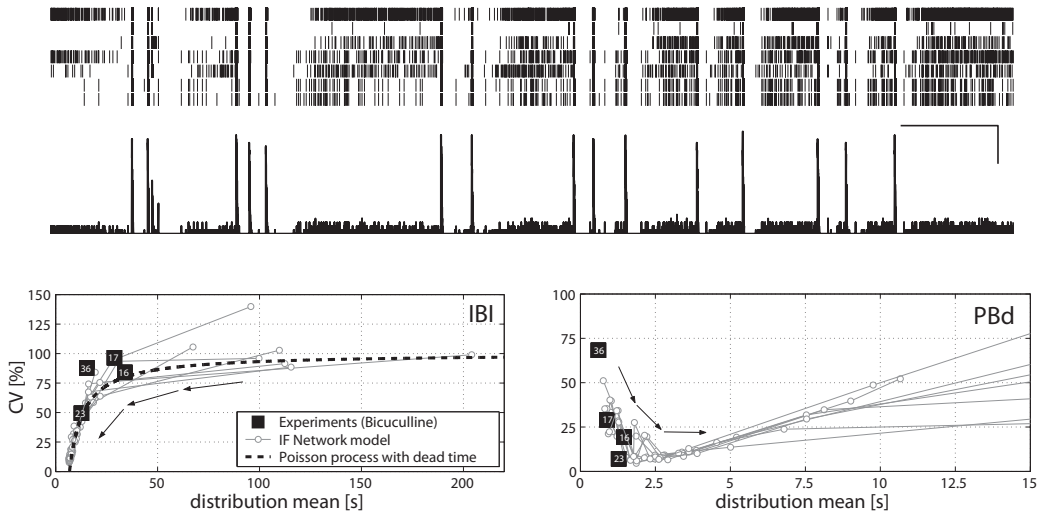
The stationary response function of the model neuron defined by Eq. 2, with spike and reset mechanisms as specified above, is by definition the output firing rate as a function of (constant)  $m_I$  and  $s_I$ , and is given by

$$\Phi(\mu, \sigma; \tau) \equiv \left[ \tau_r + \tau \int_{V_r}^{\hat{\theta}} \sqrt{\pi} e^{u^2} (1 + \text{erf}(u)) du \right]^{-1}, \quad (4)$$

where the ‘‘hat’’ operation applied to  $\theta$  and  $V_r$  is defined by  $\hat{z} \equiv (z - \mu\tau)/\sigma\sqrt{\tau}$ . In the presence of firing rate adaptation, the adapted firing rate  $f$  can be obtained as the self-consistent solution of

$$f = \Phi(m_I - \alpha f, s_I), \quad (5)$$

where the parameter  $\alpha$  quantifies the strength of adaptation. The derivation of Eq. 5 from a minimal model of firing rate adaptation, and the conditions for it to hold valid, are presented in detail in La Camera et al (2008).



**Fig. 1** Network activity emerging in dissociated cultures of neurons, detected by substrate arrays of extracellular microelectrodes (MEAs). The raster plot (upper panels) indicates the occurrence of spikes detected by 7 substrate electrodes and, below, the resulting population firing rate (scale bars: 60 sec, 10 Hz). Results from 4 MEAs experiments were compared to computer simulations of small recurrent networks of IF neurons, whose excitatory synaptic coupling was increased in the direction of the arrows. Numbers associated with each marker help identifying the same experiment in both plots, reporting the coefficient of variation versus the mean, for the distribution of the interburst intervals (IBI) as well as for the population bursts (PBd). Used and modified with permission from Giugliano et al (2004). Copyright © 2004 by the American Physiological Society.

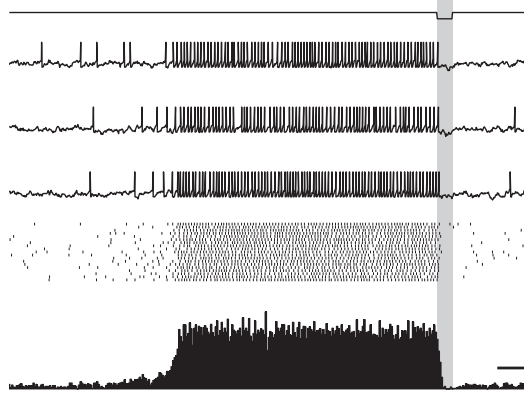
## 2 Rhythmic activity and population bursts

Cultures of neurons from rat neocortex exhibit spontaneous, temporally patterned, network activity (Fig. 1). Neurons enzymatically dissociated from embryonic (rat) neocortex can be cultured *in vitro* and maintained under healthy conditions for up to several months in an incubator (Potter and DeMarse 2001). Neurons remain electrically active (Kamioka et al 1996; Van den Pol et al 1996), mature and continue developing *ex vivo*, and spontaneously reorganize into functional synaptic networks (Nakanishi and Kukita 1998) over the 2D surface of a Petri dish, or of an array of substrate micro-electrodes (Giugliano and Martinoia 2006). As opposed to brain slices, where spontaneous activity is largely abolished by the deafferentation following acute tissue cut, neuronal cultures spontaneously display a variety of collective spiking states (Wagenaar et al 2006; Marom and Shahaf 2002). Thus, they offer a unique framework for identifying the response properties of individual neurons relevant to the collective dynamics of the whole network. In this section, we review the application of the mean field approach to the analysis of patterned rhythmic activity in those networks (Giugliano et al 2004).

### 2.1 Recurrent networks of IF neurons reproduce bursting activity

Small recurrent networks of 100 – 1000 IF excitatory neurons, incorporating the model parameters identified in the single-neuron response function experiments reviewed La Camera et al (2008) (see their Table 1), were able to capture

the spontaneous rhythmic activity of Fig. 1 (Giugliano et al 2004). To mimic the experimental conditions, where the action of inhibitory neurons is blocked by appropriate selective chemical antagonists (Giugliano et al 2006), no inhibitory input was considered in the model network. The simulated network displayed spontaneously alternating intervals of asynchronous activity at low firing rate, and bursts of action potentials synchronized across the entire population (Fig. 2). To obtain this behavior, a modest background synaptic activity, meant to parallel the spontaneous release of neurotransmitter observed in mature and immature cultures, was sufficient. Note that individual model neurons were not intrinsically bursting cells; instead, the synchronous emergence of brief epochs of intense firing rate were caused their recurrent synaptic interactions. The spatial location, from which a network-wide synchronization event originates, varied randomly, consistent with the lack of a spatial structure in the network, and in keeping with the experimental findings (Fig. 1) (Kamioka et al 1996; Giugliano et al 2004). Varying the average synaptic coupling strength and the overall number of synaptic connections mimicked different stages of *in vitro* synaptogenesis and development (Kamioka et al 1996). For very weak coupling, the model network generated no synchronous burst of activity, in agreement to what observed during the first 5-7 days after plating (Giugliano et al 2006). For larger but still weak coupling, the network generated rare and unpredictable population bursts which became more frequent and regular as soon as the excitatory synaptic coupling was increased (Fig. 1). The duration of the synchronized spiking epochs had comparable statistical properties in simulations and experiments (Giugliano et al 2004).



**Fig. 2** A recurrent network, constituted by 100 excitatory identical IF model neurons, shows a spontaneous transition from a stable resting state  $f_{rest}$  to another  $f_{burst}$  due to the finite-size fluctuations. For the sake of illustration firing rate adaptation was not included (i.e.  $\alpha = 0$  in all neurons, see Eq. 5), so that  $f_{burst}$  is a stable stationary state of the network (scale bars; 200 ms and 25 Hz/50 mV). The second transition  $f_{burst} \rightarrow f_{rest}$  is induced by transiently reducing  $m_0$  (upper continuous trace). Used and modified with permission from Giugliano et al (2004). Copyright © 2004 by the American Physiological Society.

## 2.2 Mean field analysis of slow rhythmic activity

In Fig. 2, the network flips between two states of activity either because of a strong enough finite-size fluctuation, or because of the application of a transient stimulus. However, in the presence of firing rate adaptation it is possible to obtain an alternative sequence of flips between the two stable states without the application of transient stimuli. We show here how it is possible to predict this phenomenon by using the single-neuron response function in the mean field approach.

In the mean field approximation, the total synaptic current received by a generic model neuron is Gauss-distributed, with its steady-state mean and variance given by

$$m_I(f) = cNJf\tau + m_0, \quad s_I^2(f) = cNJ^2f\tau/2 + s_0^2. \quad (6)$$

Here,  $f$  is the mean firing rate of the network,  $N$  is the number of neurons of the network,  $c$  is its “connectivity” (i.e., the average fraction of connections per neuron), and  $J$  is the amplitude of the synaptic weights (Amit and Brunel 1997).  $m_0$  and  $s_0^2$  reflect the mean and variance of the current generated by spontaneous neurotransmitter release and other sources of randomness, assumed to be independent of  $f$ . The equilibria of the population dynamics can be identified as the self-consistent solution of the mean field equation (La Camera et al 2008):

$$f = \Phi(m_I(f), s_I(f)). \quad (7)$$

The fixed points at which the slope of the response function is less than 1 are stable.

For appropriate values of  $J$ , two stable equilibria can be found that we name here  $f_{rest}$  and  $f_{burst}$ , with  $f_{rest} < f_{burst}$  (see Fig. 3A, the curve with  $J = 12$  pA). In the model network, spontaneous transitions from one of these stable states to the other can only occur due to fluctuations induced by finite-size effects (Brunel and Hakim 1999; Mattia and Del Giudice 2002), or triggered by an external stimulus (as in Fig. 2).

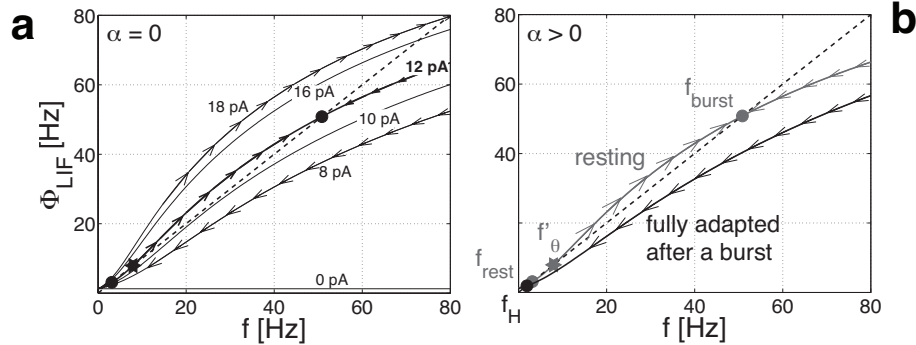
However, in the presence of firing rate adaptation, the state point  $f_{burst}$  may destabilize after the neurons undergo adaptation (Fig. 3B). To understand why this may occur, notice that in the presence of adaptation the fixed points of the network are given by the solution of the self-consistent equation

$$f = \Phi(m_I(f) - \alpha f, s_I(f)), \quad (8)$$

(see Eq. 5). When  $f$  is very small, like in state  $f_{rest}$ , the adapting term  $-\alpha f$  is also small, and the properties of this state do not change appreciably. Things are different in the  $f_{burst}$  state, however, as shown in Fig. 3B. Intuitively, while  $f_{rest}$  remains a stable solution of Eq. 8,  $f_{burst}$  is a solution only transiently, until adaptation has fully built up. This explains in simple terms the mechanisms behind the suppression of a single synchronized network event, or “population burst”, and parallels the transient depression of firing seen e.g. in Fig. 1.

## 3 Population response to time-varying inputs

In this section, we show how the single-neuron response function can be used to predict the dynamic behavior of networks of spiking neurons. This application goes beyond the stationary framework in which the mean field approach is derived, and provides us with an approximated solution of a complex Fokker-Plank equation in a two or higher dimensional space (Brunel and Sergi 1998; Brunel and Hakim 1999; Moreno et al 2002; Nykamp and Tranchina 2001; Fourcaud and Brunel 2002; Moreno-Bote and Parga 2004; Gigante et al 2007a,b). We present a case study where the use of the stationary response function provides a good approximation to the full approach (Renart et al 2003). This approximation can be extended to include firing rate adaptation, generalizing Eq. 5 (La Camera et al 2004).



**Fig. 3** Prediction of slow rhythmic activity from the properties of the single-neuron response function. Each curve is the firing rate of the population of LIF neurons ( $\Phi_{LIF}$ ) as a function of the firing rate of the population itself ( $f$ ). When  $\Phi_{LIF} = f$ , the networks has reached a self-consistent state, which may be stable (closed circles) or unstable (star), depending on the slope of the response function at  $f$ . The slope of the response can be controlled by changing the value of the synaptic couplings  $J$ , both in the absence (a) and in presence (b) of adaptation. **a.** For  $J = 8$  or  $10$  pA, only the stable state at low firing rate is stable, named here  $f_{rest}$ . For  $J = 12$  pA, a state with higher firing rate is also stable, taken here to represent a population burst (see the text). In these examples there is no firing rate adaptation ( $\alpha = 0$ , see Eq. 5). **b.** Same as (a) in the presence of firing rate adaptation (Eq. 5 with  $\Phi = \Phi_{LIF}$ ). The stability properties of the network change in the presence of adaptation: whereas  $f_{rest}$  is stable also in the presence of adaptation,  $f_{burst}$  destabilizes after the neurons undergo adaptation. Adaptation decreases the slope of the response function, which morphs into the lower curve, taking the network state into  $f_{rest}$ . In this state the neurons are negligibly adapted because of their low firing rate, and the response function becomes the upper curve again, where  $f_{burst}$  is stable. Given the small basin of attraction of state  $f_{rest}$  (this can be inferred from its distance, along the curve, from the unstable fixed point marked with '\*'), an upward fluctuation of the spontaneous activity is sufficient to bring the network back into the  $f_{burst}$  state. This state destabilizes after the neurons adapt, and the sequence of transitions repeats itself, with the network activity flipping between these two states in an activity-dependent way, as confirmed by simulations (not shown). Used and modified with permission from Giugliano et al (2004). Copyright © 2004 by the American Physiological Society.

### 3.1 Simplified mean field theory with time-varying inputs

Consider an input spike train of time-varying frequency  $\nu_x(t)$ , targeting each cell of a population of neurons through  $x$ -receptor mediated channels. Each spike contributes a post-synaptic current of the form  $\bar{g}_x e^{-t/\tau_x}$ , where  $\bar{g}_x$  is the peak conductance of the channels. In the diffusion approximation this produces an input current,  $I_x$ , which is an Ornstein-Uhlenbeck process,

$$dI_x = -\frac{I_x - \bar{m}_x}{\tau_x} dt + \bar{s}_x \xi_t \sqrt{\frac{2dt}{\tau_x}}, \quad (9)$$

with  $\bar{m}_x = \bar{g}_x \nu_x(t) \tau_x$  and  $\bar{s}_x^2(t) = (1/2) \bar{g}_x^2 \nu_x(t) \tau_x$  (La Camera et al 2008). Renart et al (2003) have suggested that the population activity of the network could be well predicted by

$$f(t) = \Phi(m_x(t), s_x^2(t)), \quad (10)$$

where  $\Phi$  is the *stationary* response function (e.g., Eq. 4), and  $m_x, s_x^2$  are the time-varying average and variance of  $I_x$ . These evolve according to the first order dynamics ( $\dot{y} \equiv dy/dt$ ):

$$\tau_x \dot{m}_x = -(m_x - \bar{m}_x) \quad (11)$$

and

$$\frac{\tau_x}{2} \dot{s}_x^2 = -(s_x^2 - \bar{s}_x^2), \quad (12)$$

(e.g., see Gardiner 1985). In this approximation, it is assumed that the network activity follows its time-varying input instantaneously. The reason for which this approximation works is that, in a network with spontaneous activity,

many neurons are close to the threshold for spike emission, making the response of the network much faster than the response of any individual neuron (e.g., van Rossum et al 2002). This justifies the assumption that the firing rate of the population is always at the steady state, despite an input with time-varying statistics  $m_x(t), s_x(t)$ .

### 3.2 Extension to networks of adapting neurons

The approach of the previous subsection can be extended to include firing rate adaptation. Consider the stationary, mean field model of the population adapted firing rate, Eq. 5, and write it as follows:

$$f = \Phi(m_x - I_\alpha, s_x^2) \quad (13)$$

$$I_\alpha = \alpha f, \quad (14)$$

where  $I_\alpha$ , called the hyperpolarization (AHP) current, is a feedback current driven by the neuron's instantaneous output rate  $f$ .  $I_\alpha$  can be derived to be of form  $-\alpha f$  from a minimal cellular model of AHP current,

$$\frac{dI_\alpha}{dt} = -\frac{I_\alpha}{\tau_\alpha} + \bar{g}_\alpha \sum_k \delta(t - t_k), \quad (15)$$

where the sum is over all spikes emitted by the neuron up to time  $t$  and  $\bar{g}_\alpha$  is the peak conductance. In the absence of actions potentials,  $I_\alpha$  decays to zero with time constant  $\tau_\alpha$ , and  $\alpha$  is linked to these cellular parameters by  $\alpha = \bar{g}_\alpha \tau_\alpha$ . This model works best in the presence of spontaneous activity (La Camera et al 2008).



From Eq. 15 one can show that, in the case of slow variation of firing rate due to adaptation,  $I_\alpha$  follows the neuron's own instantaneous firing rate with time constant  $\tau_\alpha$  (La Camera et al 2004):

$$f = \Phi(m_x - I_\alpha, s_x^2) \quad (16)$$

$$\tau_\alpha \dot{I}_\alpha = -I_\alpha + \alpha f, \quad (17)$$

Note that for a stationary stimulus, i.e.  $v_x$  constant, after a transient of the order of  $\max\{\tau_x, \tau_\alpha\}$ , one recovers the stationary model Eqs. 13-14, with  $m = \bar{m}_x$ ,  $s = \bar{s}_x$ .

In the case of several independent components, they follow their own synaptic dynamics and sum up in the argument of the response function to give the time-varying firing rate,

$$f = \Phi\left(\sum_x m_x - I_\alpha, \sum_x s_x^2\right), \quad (18)$$

which replaces Eq. 16. In Fig. 4 is shown the response of a population of uncoupled LIF neurons to a complex input, comprising an impulsive increase of 1 ms duration at  $t = 250$  ms and a step increase at  $t = 400$  ms (horizontal bars in Fig. 4) on top of the synaptic current. The synaptic current was made up of two fast components ( $\tau_x = 5$  ms), one excitatory (AMPA-like), the other inhibitory (GABA<sub>A</sub>-like), plus a third component mimicking slow (NMDA-like) current (with  $\tau_{nmda} = 100$  ms). The model makes a good prediction of the population activity, even during the fast transients. The small discrepancies are due to finite-size effects (Brunel and Hakim 1999; Mattia and Del Giudice 2002), and to the approximation used for the stationary response function (Eq. 4 corrected for fast correlated inputs (Brunel and Sergi 1998; Fourcaud and Brunel 2002)). To be able to use this approximation, the synaptic current must be much faster than the membrane time constant, i.e.,

$$\max_x\{\tau_x\} \ll \tau. \quad (19)$$

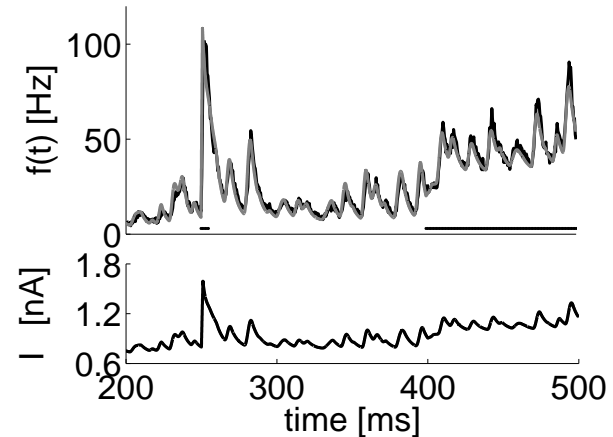
Synaptic current with longer correlation times (e.g., NMDA-like or GABA<sub>B</sub>-like) can be approximated by a slowly changing current, i.e., their variance can be neglected in Eq. 18 (Brunel and Wang 2001).

### 3.3 Further extensions

The approach of this section can be extended to any model neuron whose response function,  $\Phi$ , is known, and not just to the LIF neuron Eq. 2. In particular, it can be extended to the conductance-based LIF neuron, with the caveat that condition Eq. 19 may not be fulfilled for currents with short time constants (e.g., AMPA-like and GABA<sub>A</sub>-like). This is because the effective time constant of the conductance-based neuron,

$$\tau^* = \left(\frac{1}{\tau} + g_e v_e + g_i v_i\right)^{-1}, \quad (20)$$

where  $g_{e,i}$ ,  $v_{e,i}$  are the excitatory (e) and inhibitory (i) input conductances and firing rates respectively, can reach values as small as a few ms, depending on the instantaneous firing



**Fig. 4** Time-varying activity of a population of independent, adapting LIF neurons in response to a noisy, broadband stimulus. (Top) prediction of the adapting rate model, Eqs. 16-17 (gray), compared to the simulations of 20,000 neurons (black). Shown is the activity after a transient of 200 ms. The short horizontal bar indicates a pulse-like increase of 1ms duration in the input current. The long horizontal bar indicates a step-like increase of both the excitatory and inhibitory input current. (Bottom) Average time course of the stimulus (fluctuations around this average are not shown). See the text and La Camera et al (2004) for details. Used with permission from La Camera et al (2004). Copyright © 2004 by The MIT Press.

rates. When condition Eq. 19 is not fulfilled, the approximated response function given by Moreno-Bote and Parga (2005) should be used (see La Camera et al 2004, 2008 for details).

## 4 Adaption over multiple time-scales

The simplified dynamic mean field theory of the previous section could be extended to work in the presence of a single adaptation current of the AHP type,  $I_\alpha$ . This model of adaptation is general enough to describe most situations of interest (Benda and Herz 2003; La Camera et al 2004). However, firing rate adaptation is a complex phenomenon occurring on several time-scales and affected by different ion currents (Thorson and Biederman-Thorson 1974; Millhauser et al 1988; Lowen, S.B. and Teich, M.C. 1992; Xu et al 1996; Sah 1996; Schwindt et al 1997; Powers et al 1999; Ulanovsky et al 2004; Gilboa et al 2005; Descalzo et al 2005; Drew and Abbott 2006; La Camera et al 2006; Wark et al 2007). As a consequence, a single adapting component may not be enough to describe the time course of cortical neurons, even in response to stationary input current (La Camera et al 2006). Despite a large number of adapting processes acting on different time-scales, an IF reduction of adapting spike trains could be obtained with no more than 4 independent adapting components in the case of pyramidal and fast-spiking (FS) neurons of the rat somatosensory cortex. These adapting processes acted on time-scales ranging from a few ms to tens of seconds (Sec. 4.3). In pyramidal neurons, one of these components was found to facilitate the firing rate

(i.e., to increase it) with a time constant of the order of 1s. In this section, we review the model used to capture the temporal aspect of the instantaneous firing rate in response to fluctuating inputs with stationary statistics.

#### 4.1 The generalized IF model with multiple adapting processes

Recall that the model neuron having the quasi-stationary firing rate given by Eqs. 16-17 is described by a single variable, the membrane potential  $V$ , which below the spike threshold  $\theta$  obeys Eq. 2 with  $\mu = (m_I - I_\alpha)/C$ , and  $I_\alpha$  given by Eq. 15.

A straightforward generalization of this model can be obtained by adding two or more independent components,  $I_\alpha = \sum_k I_{\alpha_k}$ , in Eq. 16 (La Camera et al 2006). Each of the component processes obeys an equation like Eq. 15, with corresponding  $\tau_k$ ,  $\bar{g}_k$  such that  $\bar{g}_k \tau_k \equiv \alpha_k$ , and  $\sum_k \alpha_k = \alpha$  (negative  $\alpha_k$ s correspond to facilitating processes, i.e., processes increasing the firing rate over time). This constraint ensures that the quasi-stationary firing rate of each spike train (reached when  $t \gg \max_k \tau_k$ ) agrees with that given by the stationary solution in mean field, i.e., by the self-consistent solution of Eqs. 13-14.

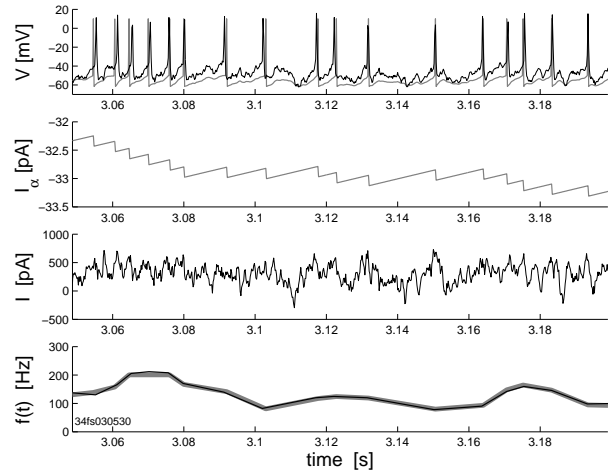
#### 4.2 Reproducing the temporal response of cortical neurons

The generalized adaptive model has been used to fit the time course of the firing rate of cortical neurons, defined as the inverse of the inter-spike interval (ISI) as a function of time,  $f(t) = 1/ISI(t)$ , using a Montecarlo procedure (La Camera et al 2006). The goodness-of-fit was then tested with  $\chi^2$  test, using as the objective function the squared difference of the ISIs, i.e.  $\chi^2_{ISI} = \sum_j (ISI_j^{exp} - ISI_j^{th})^2$ .  $ISI^{th}(t)$  was obtained by simulating the full IF model described in the previous section. Note that it is necessary to consider the correct number of processes in order to estimate the time constants involved: since  $\sum_{k=1}^n \bar{g}_k \tau_k = \alpha$ , this constraint will be satisfied with different values of  $\tau_k$  depending on  $n$ , the total number of processes. The peak conductances  $\bar{g}_k$  were given by the ratio  $\alpha_k/\tau_k$  (negative if  $\alpha_k < 0$ , representing facilitation instead of adaptation). An example showing the model with one component of adaptation and its best fit to a FS neuron of rat somatosensory cortex is shown in Fig. 5.

This procedure was not used to reproduce spike times with a millisecond precision (e.g., Jolivet et al (2004, 2006)), but only to capture the time course of the instantaneous firing rate.

#### 4.3 Time-scales of temporal adaptation in cortical neurons

In response to a stimulation of several seconds with fluctuating current with stationary statistics, the temporal response of FS interneurons of layers 2/3 and layer 5 of rat somatosensory cortex could be captured with the generalized adaptive

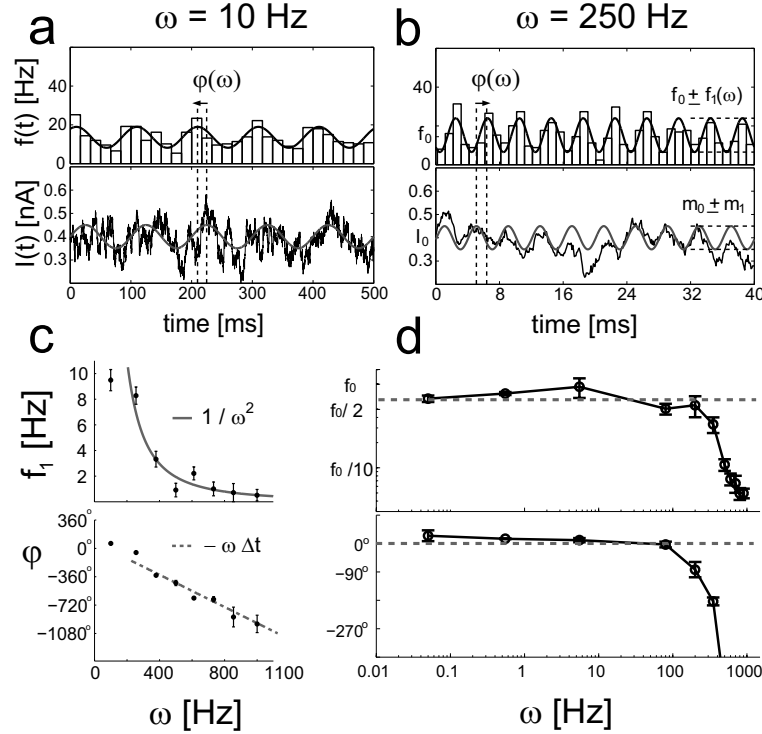


**Fig. 5** Generalized adaptive LIF model and its best fit of the temporal response of a layer 5 FS interneuron of rat somatosensory cortex. Shown (from top to bottom in each panel) are the membrane potential, the adaptation current (Eq. 15 with time constant  $\tau_\alpha = 2.2$  s), the total input current, and the instantaneous firing rate (see the text). The input current was the same as injected into the neuron. In top and bottom panels, the model is in gray and the data in black. Used and modified with permission from La Camera et al (2006). Copyright © 2006 by the American Physiological Society.

LIF neuron of Fig. 5 with only two components. The faster adapting process was of the order of  $\sim 200$ ms; the slower had a broader distribution of time constants ranging from  $\sim 1$  to  $\sim 10$  seconds (La Camera et al 2006). Given that the maximal interval duration of stimulation was 10s, it is not excluded that even slower adapting processes could be present in these neurons.

Contrary to FS neurons, which typically had a consistent response to the same stimulus, pyramidal neurons of rat somatosensory cortex display a broader range of time constants and magnitude of the adapting processes involved. Four processes were identified according to the order of magnitude of their time constants, which were, from faster to slowest: a few milliseconds (affecting the first few ISIs only), 50-200ms, 0.5-1s (a facilitatory process), and order of seconds.

Despite the possibility to capture the temporal response of cortical neurons with only a handful of adapting processes, the distributions of time constants were broad, both within cells (pyramidal neurons) and across cells (pyramidal and FS neurons; see Table 3 of La Camera et al 2006 for details). This variability in response to stimuli and across neurons suggests the possibility of a continuum of time-scales, which may be the basis for scale-free adaptation (Fairhall et al 2001b; Drew and Abbott 2006), a phenomenon with many potential computational consequences (Brenner et al 2000; Fairhall et al 2001a; Drew and Abbott 2006; Wark et al 2007). Most of these computational consequences find application in response to stimuli with time-varying statistics; however, it is important to bear in mind that multiple



**Fig. 6** Noisy sinusoidal input currents were injected in cortical neurons firing at 10-20 spikes/sec to probe their dynamic response properties (a-b) and the impact of input fluctuations. The mean input  $m_0$  was modulated as  $m_I = m_0 + m_1 \sin(2\pi\omega t)$ , with  $m_1 \ll m_0$ , while the amplitude of the background noise ( $s_I$ ) was kept constant. The neuron's output firing rate was also sinusoidally modulated at the same frequency  $\omega$ ,  $f(t) = f_0 + f_1(\omega) \sin(2\pi\omega t + \phi(\omega))$ , over a wide range of input frequencies, 0 – 200 Hz (c: single cell; d: population).  $f_0$  is the stationary response function (e.g., Eq. 4 for the case of the LIF neuron),  $f_1$  is the response amplitude of the modulated response and  $\phi$  is its phase shift. Used and modified with permission from Köndgen et al (2008). Copyright © 2008, Oxford University Press.

adaptation processes are also observed in response to inputs with stationary statistics, as reviewed in this section.

## 5 Response to sinusoidal inputs in the presence of fluctuations

The dynamic mean field theory of the previous sections is a simplified approach which, in those cases where it can be applied, offers practical advantages to the study of the temporal evolution of the network activity in the presence of time-varying inputs. In the general case, the full population density approach should be used, but the equations are complex and are usually solved with perturbative techniques for the case of weak (but arbitrarily fast) input modulation. In this section, we reviewed some of the results obtained with this approach, which dates back to Knight (1972); Treves (1993) (in the absence of noise), was generalized by Gerstner (2000), and has more recently been undertaken by Brunel and collaborators (Brunel et al 2001; Fourcaud and Brunel 2002; Fourcaud-Trocmé et al 2003; Fourcaud-Trocmé and Brunel 2005; see also Mattia and Del Giudice 2002).

The idea behind this approach is to analyze the response of the network to a weak oscillatory input (i.e., sinusoidal with frequency  $\omega$ ),

$$m_I = m_0 + m_1 \sin(2\pi\omega t), \quad (21)$$

with  $m_1 \ll m_0$ , in the presence of input fluctuations with constant amplitude  $s_I$  (see e.g. Fig. 6a-b). The response of the network to this input can be found by solving perturbatively a Fokker-Planck equation in series of  $1/\omega$ , assuming that the neuron's output firing rate is also sinusoidally modulated at the same frequency,

$$f(t) = f_0 + f_1(\omega) \sin(2\pi\omega t + \phi(\omega)). \quad (22)$$

In networks of LIF neurons with instantaneous synapses, the amplitude of the response modulation ( $f_1(\omega)$ ) in the high-frequency regime is proportional to  $1/\sqrt{\omega}$ , with a phase lag of 45 degrees (Brunel et al 2001). In the presence of synaptic filtering, both the cut-off frequency and the phase lag are eliminated (Brunel et al 2001). However, another important determinant to the response to oscillatory inputs, the mechanism for spike generation, is absent in the LIF neuron. For this reason, generalized models of IF neurons, where a fast transient response of the membrane potential above a given threshold can mimic an intrinsic mechanism of spike generation, were introduced and studied by Fourcaud-Trocmé

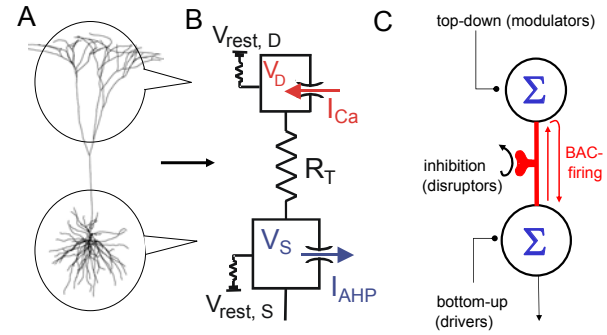


et al (2003) (called nonlinear IF models). These models are obtained by adding to the right hand side of Eq. 2 a term  $\psi(V)/C$ , which contains the non-linear mechanism underlying the generation of an action potential, and a parameter  $\Delta_T$  defining its sharpness. The firing rate modulation at high frequencies depends on  $\psi$ , being e.g. of the form  $1/\omega$  for an exponential function, and  $1/\omega^2$  for the quadratic case (Fourcaud-Trocmé et al 2003). These results are also valid in the presence of conductance-based synaptic inputs, and suggest that in general the high-frequency modulation of the response follows a power law  $\omega^{-\gamma}$ , where the exponent  $\gamma$  depends on the nonlinearity of the spike generating current  $\psi(V)$  (Fourcaud-Trocmé et al 2003). The (unmodulated) background synaptic noise also plays a role in the determination of the linear response (Brunel et al 2001; Fourcaud and Brunel 2002), and so does the presence of firing rate adaptation (Fuhrmann et al 2002).

These theoretical results provide a number of precise predictions which were tested experimentally by Köndgen et al (2008) in rat cortical pyramidal neurons of the somatosensory cortex (Fig. 6). These authors found that the amplitude of the linear modulation ( $f_1(\omega)$ ) was generally constant and independent of  $\omega$  up to rather high input frequencies, below a sharp cut-off of the order of 100 – 200 Hz. In this range, no phase-shift was observed other than a phase-advance, caused by spike-frequency adaptation at frequencies lower than 10 Hz (Fuhrmann et al 2002) (Fig. 6c-d). This confirmed the role of the background synaptic noise in removing the resonances at multiples of the average firing rate response  $f_0$  (Brunel et al 2001; Fourcaud and Brunel 2002), allowing fast time-varying inputs to be encoded undistorted (i.e., without a phase lag). Beyond the cut-off frequency range, the high frequency linear response was found to decay as  $f_1(\omega) \sim \omega^{-\gamma}$ , with  $\gamma > 1$  and close to 2, independently of the noise correlation time constant (Fig. 6c-d). This behavior is consistent with a nonlinear IF model with a very sharp spike, and a nonlinearity which is intermediate between exponential and quadratic (Fourcaud-Trocmé and Brunel 2005).

## 6 Response to dendritic inputs and soma-dendritic interactions

So far, we have been concerned with those properties of single neurons and network activity that could be analyzed assuming a point-neuron model. Point-neuron approximations may be a good description for small neurons with short and isotropic dendritic trees. However, the apical dendritic tree of layer 5 pyramidal neurons extends across all cortical layers with a length of roughly 1.5 mm, and integrate inputs from different cortical and subcortical sources (Budd 1998; Binzegger et al 2004; Oda et al 2004). Whether the extended geometry of pyramidal neurons offers real computational advantages, or whether it only solves the ‘packing problem’ of collecting a large amount of synapses for a single integration process, remains an open issue and it will



**Fig. 7** 2-compartmental model of a cortical L5 pyramidal neuron. A) Beside the basal integration zone around the soma (lower circle) L5 pyramidal neurons show an additional apical integration zone (upper circle) from where the signaling to the soma is mediated via dendritic calcium spike. (B) To capture the spatial input structure we consider a 2-compartment neuron model with a voltage-dependent calcium current ( $I_{Ca}$ ) in the dendritic compartment and a spike-triggered after-hyperpolarization current ( $I_{AHP}$ ) in the somatic compartment. (C) Dendritic (‘top-down’) input increases the gain of the somatically (‘bottom-up’) induced current-to-rate response function via backpropagating action potential induced calcium (BAC) firing. This top-down input represents a multiplicative modulation of the somatic firing rate (with a factor  $\geq 1$ ) which can be throttled by inhibitory dendritic input. Hence, the dendritic tree determines whether synaptic input either acts as ‘driver’, ‘modulator’, or ‘disruptor’ (Sherman and Guillery 1998), depending on whether it projects to the somatic region or the apical tuft, and whether it acts through excitation or inhibition.

not be discussed here. Instead, we will focus on some of the phenomena that depend critically on such extended geometry and cannot be captured with point-neuron approximations, like the dendritically-induced gain modulation of the somatic response and its control by inhibition (Larkum et al 2004; Murayama et al 2008). This will allow us to characterize the response of layer 5 pyramidal neurons to noisy input currents which are simultaneously injected in the soma and in the apical dendrite.

### 6.1 Dendrites can differentiate between drivers, modulators and disruptors

To structure the possible functional interactions among neurons, it has been suggested to classify neurons into drivers, modulators, and disruptors (Sherman and Guillery 1998). While drivers act additively on the output frequency, modulators act multiplicatively and disruptors block either of them. The multiplicative scaling of the neuronal response function is an efficient mechanism for modulating the neuronal responses in one cortical area by the activity in another area, which may be an important mechanism for cognitive processing (Salinas and Thier 2000).

It has been noted that a point neuron is able to integrate both drivers and modulators with the same type of ionotropic synapses (Chance et al 2002; Abbott and Chance 2005). Here we show that an extended dendritic tree allows

to incorporate all three operations by varying the location of the synaptic inputs (Fig. 7).

Experimental data and modeling studies confirm that excitatory synaptic input projecting to the proximal dendritic tree can act efficiently as a driver (i.e., it shifts the response function to the left), while synaptic input to the distal dendritic tree can also act as a modulator (i.e., it also increases the gain of the response function), see Fig. 8A and (Larkum et al 2004). The basic mechanism allowing the distal dendritic input to modulate the gain relies on the generation of dendritic calcium spikes. These calcium spikes may be triggered by synaptic inputs on the distal dendritic tree impinging on a back-propagating action potential (AP). The calcium-induced dendritic depolarization propagates forward to the soma where it triggers one or several additional APs. Because this mechanism generates two or more APs out of one single AP, it represents a multiplicative operation on the response function. Since it is triggered by the joint emission of a back-propagating AP and a dendritic calcium spike, this mechanism is referred to as backpropagation-induced calcium (BAC) firing (see also Larkum et al 1999).

In the experimental data, the multiplicative modulation is also accompanied by a left-shift of the response function, which otherwise characterizes the action of drivers (Fig. 7). While in the experiment the left-shift arises from a rather proximal positioning of the dendritic electrode, distal synaptic input further away from the soma will undergo a stronger attenuation and will barely contribute to the direct somatic depolarization. However, it will still contribute to the generation of a calcium spike in the apical dendrite and thus to a gain modulation of the somatic current-to-frequency response function.

Beside the action of somatic and dendritic excitation as drivers and modulators, respectively, inhibitory synaptic inputs efficiently operates as disrupters of both drivers and modulators. Inhibition may shunt the somatic voltage when the excitatory synaptic input projects proximally to the soma (Doiron et al 2001; Mitchell and Silver 2003), or it may transiently block the calcium conductance and thus disrupt the gain modulation due to calcium spikes when the excitatory synaptic input projects to the distal dendrite (Larkum et al 1999; Pérez-Garci et al 2006).

## 6.2 Two-compartmental IF model with dendritic calcium spikes

The phenomena described in the previous subsection can be explained by an extension of the LIF point-neuron model to include two compartments, representing the somatic and the distal dendritic regions, respectively (Fig. 7A,B). A successful model of the calcium-induced dendritic depolarization requires the integration of some active dendritic current ( $I_{Ca}$ , Fig. 8C). To capture the fact that a dendritic calcium spike is only triggered by a fast voltage up-sweep across some voltage interval, a dynamic activation and inactivation of the calcium conductance must be considered (this can be done in

terms of activation and inactivation variables,  $m$  and  $h$ , respectively, see Larkum et al (2004)).

Because in the experiment the calcium-induced somatic AP bursts often cease even when the dendritic membrane is still depolarized, a potassium AHP current ( $I_{AHP}$ , see Eq. 15) was included in the somatic compartment (Figs 7B and 8B,C). This transiently activated leak current represents a sort of security valve, as it prevents the model neuron from being trapped in a calcium-induced depolarization plateau where it would continuously burst.

The two-compartmental IF model reproduces the firing rates of L5 pyramidal neurons in response to somatic and dendritic current injections (Fig. 8A). It also reproduces the increased gain of the somatic current-to-frequency curve in the presence of a dendritic background input (Fig. 8A and Larkum et al (2004)). When injecting a noisy somatic input current only, the individual spikes can be well predicted (Fig. 8B). However, when injecting a dendritic input currents, it is just the strong dendritic voltage deflection ('calcium spike') and the induced AP burst which can be predicted, but not the individual spike times within the burst (Fig. 8C). A detailed description of the 2-compartmental model is provided in Larkum et al (2004).

## 6.3 Somatic response function for joint somatic and dendritic inputs

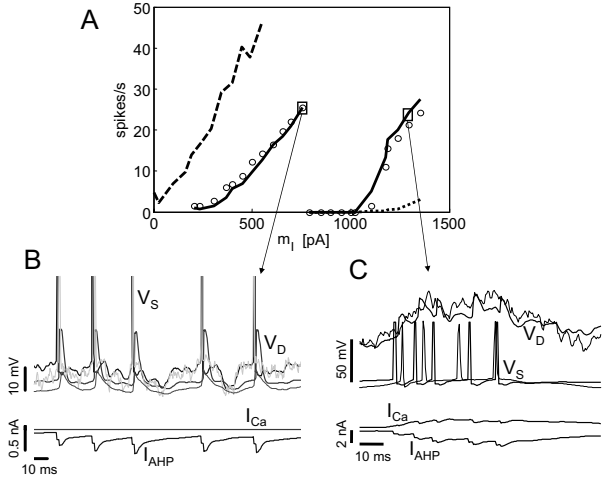
In this section we provide an approximated formula for the somatic response function of the 2-compartmental IF model of the previous sub-section. We start from an approximation to the response function Eq. 4 for a single compartment IF neuron (Abbott and Chance 2005),

$$f^{(1)} = \frac{(\bar{V} - \theta)}{\tau(\theta - V_{reset}) \left(1 - e^{-a(\bar{V} - \theta)/\sigma_V}\right)}, \quad (23)$$

where  $\tau = RC$  is the membrane time constant. Here,  $\bar{V}$  is the average of the membrane potential when the spiking generation mechanisms are inactivated, and  $\sigma_V$  is its standard deviation. These parameters are proportional to the mean and standard deviation of the input current,  $m_I$  and  $s_I$ , by a factor  $R$ . When currents of the Ornstein-Uhlenbeck type (Eq. 9) are injected both into the soma and the distal dendrites, the corresponding somatic and dendritic voltages are characterized by  $\bar{V}_S$ ,  $\sigma_{V_S}$  and  $\bar{V}_D$ ,  $\sigma_{V_D}$ , respectively.

Recall that the induction of a dendritic calcium spike requires a fast dendritic voltage sweep crossing a certain voltage threshold. It is possible to trigger a calcium spike by dendritic input currents only, but in the presence of a back-propagating AP far less dendritic input is needed. We therefore concentrate on those calcium spikes which are triggered by the joint action of a back-propagating AP and a simultaneous dendritic voltage sweep. This will lead directly to a gain modulation of the somatic current-to-frequency response function (see Eq. 26 below).

The specific requirement on the voltage transient arises from the narrow window formed by the voltage-dependent



**Fig. 8** Somatic and dendritic response functions and voltage traces for the 2-compartmental model. A) Firing rates of the model neuron in response to noisy input currents ( $s_I = 300$  pA) of varying mean ( $m_I$ ) injected individually in the soma (line starting around  $m_I = 250$  pA; circles representing experimental data) and the dendritic compartment (line with onset around  $m_I = 1000$  pA). When applying the same somatic currents in the presence of a noisy dendritic input (fixed to  $m_I = 750$  pA and  $s_I = 300$  pA) the gain of the neuronal response function with respect to the somatic input is increased (dashed curve). Blocking the calcium current in the dendrite would lead to a rather shallow response function for the dendritic current injection (dotted curve). B) The somatic ( $V_S$ ) and dendritic ( $V_D$ ) model voltage trace in the case of pure somatic current injection closely predicts the experimental spike times. The thin smooth and noisy curves show the corresponding experimental traces. The lower panel confirms that no dendritic calcium current ( $I_{Ca}$ ) is elicited in the model, whereas a strong after-hyperpolarization current ( $I_{AHP}$ ) is triggered after each spike. C) In the presence of a dendritic input, the dendritic calcium current ( $I_{Ca}$ ) is responsible for the strong dendritic depolarization ( $V_D$ ) generating the ‘BAC-burst’ ( $V_S$ ). The parameters used in these simulations were (see Sec. 1.1 for a definition of the symbols, with ‘S’ standing for somatic compartment, and ‘D’ standing for dendritic compartment):  $R_S = R_D = 45$  mV,  $R_T = 70$  mV,  $V_{rest,S} = -70$  mV,  $V_{rest,D} = -60$  mV,  $C_S = \tau_S/R_S = 13/45$  nF,  $C_D = 5/45$  nF. The AHP current was as in Eq. 15, with the inclusion of a reversal potential of  $-90$  mV and  $g_\alpha = 5$  nS,  $\tau_\alpha = 60$  ms. The calcium current ( $I_{Ca}$ , see Larkum et al 2004 for details) is specified by  $g_{Ca} = 70$  nS,  $E_{Ca} = 120$  mV,  $m_{1/2}^\infty = 0$  mV,  $h_{1/2}^\infty = -10$  mV,  $\text{slope}(m^\infty) = 1/3$  (mV) $^{-1}$ ,  $\text{slope}(h^\infty) = 1/5$  (mV) $^{-1}$ ,  $\tau_m = 15$  ms and  $\tau_h = 80$  ms. A somatic AP is elicited when  $V_S$  crosses the threshold  $\theta = -47$  mV and this activates an additional  $I_{AHP}$  component. Subsequently,  $V_S$  is clamped for 1 ms at 20 mV and then reset for another 1 ms at  $-52$  mV, right 5 mV below the threshold. To mimic the back-propagating AP, the dendritic voltage  $V_D$  is instantaneously raised by 20 mV with a delay of 4 ms after  $V_S$  crossed the threshold.

activation and inactivation functions ( $m^\infty$  and  $h^\infty$ , respectively) which are typical for calcium currents (Koch 1999; Larkum et al 2004). Since only a weak steady state window current is possible with these gating functions, a slowly increasing dendritic voltage would merely inactivate the calcium conductance without being able to trigger a calcium spike. Dendritic voltage traces produced by an Ornstein-Uhlenbeck process will therefore only lead to calcium spikes if the membrane potential deflections (characterized by  $\sigma_{V_D}$ ) are large, and especially if these deflections arise together

with a dendritic steady state depolarization ( $\bar{V}_D$ ) which is strong compared to the AP threshold ( $\theta$ ). Hence, a criterium for the generation of a dendritic calcium spike given a somatically induced AP may have the form

$$\sigma_{V_D}(1 + \bar{V}_D/\theta) \geq \theta, \quad (24)$$

where for the sake of simplicity we have chosen the same threshold  $\theta$  for the generation of a calcium spike as for the generation of an AP. The probability,  $P_{Ca}$ , of a dendritic calcium spike, conditioned on the earlier occurrence of a somatic AP within a short time interval, is a saturating, increasing function of the  $V_D/\theta$  term in Eq. 24; it can be expressed as

$$P_{Ca} = \left(1 - e^{-[\sigma_{V_D}(1 + \bar{V}_D/\theta) - \theta]/b}\right) \rho \quad (25)$$

where  $b$  is a positive constant in units of voltage and  $\rho$  is a scaling factor between 0 and 1.

Next we consider the impact of the dendritic calcium spikes on the neuronal response function. The firing rate  $f^{(2)}$  of the 2-compartment model is due to the APs generated by the somatic current injection and the additional APs generated by BAC-firing. Each AP induces an iterative, albeit short-lived, process: a dendritic calcium spike occurring with probability  $P_{Ca}$  causes, on average, a subsequent AP at the soma, which in turn has a probability  $P_{Ca}$  to generate another AP due to a calcium spike, and so on.  $f^{(2)}$  can then be obtained as the sum across all iteratively induced APs, starting with basic firing rate  $f^{(1)}$  of the single compartment,

$$f^{(2)} = f^{(1)} \sum_{i=0}^{\infty} (P_{Ca})^i = \frac{f^{(1)}}{1 - P_{Ca}}, \quad (26)$$

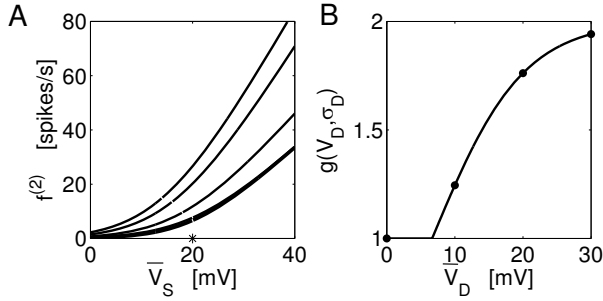
where the right-hand-side is obtained by summing the geometric series. Alternatively, one may obtain  $f^{(2)}$  from solving the recursion relation  $f^{(2)} = f^{(1)} + f^{(2)}P_{Ca}$ . Plugging the expressions for  $f^{(2)}$  and  $P_{Ca}$  (Eqs 23 and 25, respectively) into the right-hand side of Eq. 26, one obtains the firing rate of the 2-compartment IF model (see Fig. 9),

$$f^{(2)} = \frac{g(\bar{V}_D, \sigma_D) (\bar{V}_S - \theta)}{\tau(\theta - V_{reset}) \left(1 - e^{-a(\bar{V}_S - \theta)/\sigma_{V_S}}\right)}, \quad \text{with} \quad (27)$$

$$g(\bar{V}_D, \sigma_D) = \frac{1}{(1 - \rho) + \rho e^{-[\sigma_{V_D}(1 + \bar{V}_D/\theta) - \theta]/b}}. \quad (28)$$

Expression (27) generalizes the 1-compartmental response function (23) to the case of two compartments, where the 2nd dendritic compartment acts as a gain modulator. The somatic and dendritic voltage variables  $\bar{V}_S$ ,  $\sigma_{V_S}$  and  $\bar{V}_D$ ,  $\sigma_{V_D}$ , respectively, are related to the somatically and dendritically injected Ornstein-Uhlenbeck currents.

The 2-compartment response function (27) neglects the passive propagation of the dendritic voltage towards the soma, and of the somatic voltage towards the dendrites. A passive dendro-somatic attenuation could be included into the model by substituting  $V_S \rightarrow V_S + \alpha_D V_D$  and  $\sigma_S \rightarrow \sigma_S + \alpha_D \sigma_D$ , with  $\alpha_D$  representing a dendritic attenuation factor. The passive



**Fig. 9** Somatic response function for different dendritic inputs in the approximation of Sec. 6.3. A) Firing rate  $f^{(2)}$  for the 2-compartment model (Eq. 27) as a function of the average somatic depolarization  $\bar{V}_S$ , plotted for different values of the average dendritic depolarizations ( $\bar{V}_D = 0, 10, 20, 30$  mV from bottom to top, with fixed  $\sigma_{V_S} = 20$  mV and  $\sigma_{V_D} = 15$  mV). The star represents the voltage threshold  $\theta = 20$  mV. The lowest curve, representing the case  $\bar{V}_D = 0$  (but with  $\sigma_{V_D} = 15$  mV), almost coincides with curve for pure somatic injection (Eq. 23). B) The gain  $g$  of the response functions in A as a function of average dendritic depolarization  $\bar{V}_D$  in the presence of a fixed noise amplitude  $\sigma_{V_D} = 15$  mV (Eq. 28). The dots specify the values for  $\bar{V}_D$  and  $g$  used to obtain the 4 curves in A. Other parameter values:  $a = 5$ ,  $b = 5$  mV,  $\rho = 0.5$ ,  $\alpha = 0.2$ ,  $\tau = 30$  ms.

component would cause a left-shift in the response function, whose strength is quantified by  $\alpha_D$ . In the experiments, this factor appeared to be relatively large ( $\sim 0.4$ ). However, since in the experiments the dendritic electrode did not reach the calcium triggering zone at the apical bifurcation (Larkum et al 2004), this factor may be much smaller in reality. Thus, the left-shift of the response function induced by the passive dendritic input may be negligible compared to the induced gain increase (cf. Fig. 9A).

## 7 Discussion

Large populations of neurons have a large number of degrees of freedom, giving rise to very rich and complex collective dynamics. In the network models reviewed in this article, the study of such a rich behavior can be simplified, because it can be reduced to the study of the time development of the distribution of the variables that characterize the single-neuron dynamics (e.g., the depolarization in the case of IF models). However, even with such a simplification, the equations governing the network dynamics are still difficult to solve. For example, in the case of weak enough synaptic interactions, the equation governing the distribution of the depolarizations of IF neurons is a Fokker-Planck equation. Although the dimensionality of the equation is low (the only dynamical variable is the depolarization), the boundary conditions, corresponding to the threshold for emitting a spike and to the lower bound of the depolarization, make it extremely difficult to find a general analytical solution. The only exact solutions known can be obtained under the assumption that the statistics of the synaptic input are stationary (La Camera et al 2008). For simple IF models, how-

ever, it is possible to simplify further the study of the time-dependent network dynamics in at least two situations: 1) when the relaxation dynamics of the Fokker-Planck equation are significantly faster than the time scale over which the input varies (as in the case of slow synaptic currents), and 2) when the amplitude of the modulations of the input statistics are small (perturbation theory). In both cases, we have shown here that it is possible to reduce the study of neural populations to the analysis of the response of single neurons to noisy inputs. This approach is possible also in the case of real neurons, for which the responses can be measured in experiments.

What could these experiments reveal that was not known for the neuronal model used to develop the theory? The experiments provided us with an estimate of the parameters of the neuron model, which is crucial for quantitative modeling studies. However, there are at least two other important, unexpected results that emerged from the analysis of the experimental measurements and that turned out to be independent of the specific neuronal model that was used to design the experiment itself. The first result is that populations of real neurons respond fast (Köndgen et al 2008), significantly faster not only than any single neuron (which was expected), but also faster than predicted by simple IF models. The linear response to sinusoidal currents is not attenuated up to frequencies of a few hundreds Hz. The second unexpected result is related to adaptation on multiple time-scales. A quantitative analysis of the response of rat pyramidal and fast-spiking neurons to long lasting, noisy stimuli with stationary statistics revealed that the activity of cortical neurons is modulated over multiple time-scales ranging from hundreds of milliseconds to seconds. The response of the neurons could be modeled with IF neurons with multiple mechanisms of adaptation and facilitation. Every neuron had up to 4 mechanisms operating on different time-scales. Moreover, the time-scales varied widely from neuron to neuron, allowing a population to show responses on almost a continuum of time-scales.

In the second part of this review, we have considered the single-neuron response to inputs that are distributed on the dendritic tree. The simplicity of the IF model contrasts with the complexity of the dendritic arborization of some pyramidal neurons, with their regenerative membrane currents and clustered synaptic inputs (Spruston 2008). The extended geometry of cortical neurons and their non-linear dendritic properties may offer additional computational power by exploiting nonlinear dendritic properties (Poirazi et al 2003; Polsky et al 2004), like the multiplicative gain modulation of the somatic response function reviewed here.

A nonlinear modulation of the response function may originate from many mechanisms, e.g., could be controlled by neuromodulators (Zhang and Arsenault 2005; Thurley et al 2008), or by the strength of after-hyperpolarization currents (Higgs et al 2006), or through a balanced change of noisy excitation and inhibition (Doiron et al 2001; Chance et al 2002; Longtin et al 2002; Mitchell and Silver 2003). These forms of gain modulation are implementable in point

neurons, and were not considered here. Other forms require instead multi-compartmentalization, like those arising from passive dendritic integration (Prescott and De Koninck 2003; Mehaffey et al 2005; Capaday and van Vreeswijk 2006).

The form reviewed here in some detail is an active form of dendritically-controlled gain modulation, i.e., mediated by BAC firing and dendritic calcium spikes. This phenomenon relies on regenerative dendritic calcium conductances and can be tuned by acting directly on those conductances (Larkum et al 2004), e.g. by selectively blocking them through activation of dendritic GABA<sub>A</sub> and GABA<sub>B</sub> receptors (Pérez-Garci et al 2006). These tuning options extend the functionality of pyramidal neurons by allowing them to distinguish between synaptic drivers, modulators and disruptors (see Sherman and Guillery 1998 and Fig. 7C), and recent experimental evidence from awake rats suggests that these different types of inputs may also play a functional role *in vivo* (Murayama et al 2008).

The mechanism of BAC-firing and BAC-firing induced gain modulation is related to a type of neural code, burst-timing code, which is different from both firing rate codes (e.g., of Fig. 4 and 8A) and spike-timing codes. A calcium spike, triggered by the joint occurrence of a back-propagating AP and the dendritic input, elicits in turn several consecutive sodium spikes at the soma within roughly 30ms (Larkum et al 1999). This mechanism allows signalling to downstream neurons that some events have occurred in coincidence. Since inputs to the soma and the apical tree may originate from different cortical sources (Budd 1998; Oda et al 2004), burst-timing could provide a way to detect and signal the coincident occurrence of bottom-up and top-down signals.

For us, the ability of simplified models to capture much of the rich and varied experimental phenomenology of cortical neurons embedded in an *in vivo*-like environment, is an indication of the success of the reductionist approach in Neurophysiology. We also hope that recent observations, like gain modulation by distal dendritic inputs or the divergence of the response functions in prefrontal cortex neurons (Arsiero et al 2007), can open the door for new quantitative models and their application to analysis of network behavior. While such an interaction between theory and experiment is a widely consolidated tradition in Physics, it is becoming only slowly established in Neuroscience.

**Acknowledgements** We thank M. Arsiero, P. Darbon, H. Köndgen, M.E. Larkum, H.-R. Lüscher and A. Rauch for their many contributions to the experimental characterization of the response function of neocortical neurons. Many of the ideas and the vision behind the work described in this article were pioneered by Prof. Daniel Amit. This work is dedicated to his memory.

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