Radiation based Microscopy

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Outline

- conventional optical microscopy
- electron microscopy
- Near Field optical microscopy
- comparison with other "indirect" microscopies

Microscopy

object:

 visual description of nature
 "At a microscope <u>the surface of objects</u> is seen, magnified and made clearer, yet do <u>not</u> think that you're seeing <u>their intimate essence</u>", Feng-shen Yin-Te (1771-1810)

medicine, biology (cells, bacteria, virus, colloidal particles, ...)
 <u>applications:</u>
 geology (crystals, ...)

- material science (phases, particles,
- chemistry, physics (molecules, atoms, ...)

after the telescope (Galileo, early 1600), Anton van Leeuwenhoek (~mid 1600):

- protozoa
- spermatozoa
- bacteria
- Red blood cells

optical "microscope"

"Power" of a microscope



• magnification (linear): $A \rightarrow B = MxA$

• resolution (resolving power): capability to separately identify 2 different points (objects)

(human eye: 100-200 μm)

• enhancing techniques: Image Analysis & Processing

Diffraction and interference

Huygens principle:

Х

>

Х

X

Х

X X X



i.e. identical in-phase coherent oscillators (radiating antennas)

IN - 1 stain 0

d sin f

Rayleigh resolution limit





$$NA = n \sin \alpha$$



N.B., resolution :

- undefined for an isolated object
- ≠ magnification M

Resolution limit: wave nature of light

Nyquist sampling theorem:

 need at least 2 data points
 (i.e.1 max & 1 min) per 1 period (wavelength) to map an oscillating signal (light wave) without aliasing



(based on Fourier transform theory)

look out : *aliasing* gives *pseudo-resolution* !

Resolution limit: particle nature of light

e ^{i k}z z

Heisenberg principle : $\Delta x \Delta k_x, \ \Delta y \Delta k_y, \ \Delta z \Delta k_z > f$ if $\Delta x, \ \Delta y < \lambda, \ \Delta k_x, \ \Delta k_y$ large invariance of $k^2 = k_x^2 + k_y^2 + k_z^2$ $\Rightarrow k_z^2 < 0, \ k_z = i | k_z |$



→ Evanescent field :

spatial frequencies >1/λ decay exponentially with distance

Sensitivity of a microscope

- necessary for resolution
- not sufficient !
- e.g.: fluorescence microscopy :

fluorescent molecule dyes (F or Cr based *fluorochromes*) chemically stick (due to pH, Ca²⁺, Mg²⁺, hydrophobic/philic...) to objects of interest and label them...



rabbit nerve cells

down to "molecular detection"! ...

van Hulst JCP 18, 7799 '00



diC18 molecule spots (with polarized light)

.... but this does not mean "molecular resolution"

birth of Electron Microscopy

- Thomson 1897: discovery of the e⁻ • De Broglie 1924: $\lambda = h/p$ • Bush 1926: **E**, **B** axial fields: slow for charged particles
- Ernst Ruska 1934: TEM
 - (commercial: 1939, Siemens & Halske)
- Knoll-Von Ardenne 1935-38: TEM + scan coils
- Zworykin-Hillier-Snyder '40s SEM
- Oatley '50s-'60s

(commercial: 1965, Cambridge Instruments)



intermediate column components



TEM







SEM





- + easy to use/understand (optical image quality)
- + no thin specimens
- + high resolution, high field depth, analytical capability ~ TEM, at lower price: 200-500 k€
 → > 20'000 SEM installed worldwide, +1 / day
- slow

Why higher resolution?

$$\lambda_{\text{DeBroglie}} = h / p$$

Planck constant: h=6.63 10⁻³⁴ Js

tennis ball: M~100 g, v~100 km/h → λ_M~2 10⁻²⁴ Å
continuous (non-oscillating) wave
e⁻: m_e~9 10⁻³¹ kg, v~1/3 c → λ_e~0.02 Å ~10²² λ_M, "observable"

$$E_{\gamma} = hv = hc/\lambda_{\gamma} \xrightarrow{\gamma} \frac{\gamma}{1240 / E}$$

$$E_{e} = p_{e}^{2}/2m_{e} = h^{2}/2m_{e}\lambda_{e}^{2} \xrightarrow{\gamma} e^{1.23 / \sqrt{E}}$$

i.e. electrons – vs – photons

 \rightarrow "softer" diffraction limit

- possible radiation damage
- but: controlled potential (i.e. conductive) samples required
 - UHV environment required



Figure 4.53 Transmission electron micrograph of a NbO particle located at a grain boundary in polycrystalline alumina. Phase contrast (lattice fringes) and mass-thickness contrast vary from the alumina grain to the NbO grain



Figure 4.54 Lattice image of a rhombohedral twin in alumina

examples of **HR-TEM**:

E>=100 kV



Figure 4.55 Lattice image of a SiC particle located within an alumina grain. The alumina lies along a low-index-zone axis, and is the source of the lattice image. A moiré pattern appears within the SiC particle due to overlap between the alumina and SiC (in the direction of the electron beam)

extremely detailed informations about thin crystalline samples (after interpretation of data and for flat sample slices only)

More electron microscopes...

STEM ...

- in 1991 true atomic resolution:
 - 1.4 Angstroem
- today almost all TEM are STEM

... ESEM

1988 ElectroScan (USA)
 → Philips → FEI

T=0 °C, P=6.5-13 mbar, RH up to 100%



samples can be: - biologic (i.e. soft, dirty, degassing)- insulators (water vapor ions drain charge away)

Scan-enhanced optical microscopy



Flurophores

Focal Plane

- optical resolution improved, particularly axial
- SPM: not true 3-D (+ perturbative)

Two (or Multi) Photon Excitation

Tryptophan Multiphoton Absorption

- + not a real confocal (no pinhole) yet intrinsic confocal-like effect
 → reduced photobleaching
- + infrared exciting photons: lower damage to biological sample



needs hi excitation photon flux for near-simultaneous
2-photon emission
→ expensive pulsed laser
e.g. fs Ti:sapphire

Excitation Photobleaching Patterns



SEM electron source

column → gun (e source) + e path "cylinder"

console (control hw – sw)



electron-solid interactions



alumina

3.5

Interaction zone



- at the boundary E~kT
- as the path length in the sample increases, <E> decreases and E spread ∆E increases

- from deeper regions than SE
 → lower resolution
- BSE : low collection efficiency (<5%)
 - strong direction memory
 → shadows



Source performance parameters



"Aberrations"

→ effective probe size: $d^2 \rightarrow d'^2 = d^2 + \sum_i d_i^2$



consequences on imaging properties



SE detection

SE weak \rightarrow easy to collect at low detector bias



annular ring (in-lens) Everhart-Thornley detector

- hi inner bias accelate e- after entrance
- Faraday cage <u>screens</u> the beam



BSE imaging

Z contrast
dominant oneescape depth $R_{BS} \propto (A/Z^{0.9}) (E^{1.7}/\rho)$
for E>5 keV $\eta \sim -0.025 + 0.016 Z$ $Z \eta$
C : 6 0.05
Au: 79 0.5

Electron Channeling Pattern





e.g.: SEM of EC-STM tips

standard insulating coatings [$A_{lree} < (10 \ \mu m)^2$] :

- poliacrylics
- ероху
- Apiezon wax



Tip coating device assembly



Apiezon wax coated Au tip. The very tip (several μ m) remains uncoated.



NFO laws can be quite different from the FFO ones

NF - vs - FF example: electric dipole



 $E_{R} = 2 \left[\frac{p \cos \theta}{R^{3}} \right] (1 + i \, k \, R) \qquad e^{i \, k \, R}$ $E_{\theta} = \left[\frac{p \sin \theta}{R^{3}} \right] [1 + i \, k \, R - (k \, R)^{2}] e^{i \, k \, R}$ $H_{\phi} = \left[\frac{p \sin \theta}{R^{3}} \right] \left[-i \, k \, R - (k \, R)^{2} \right] e^{i \, k \, R}$

p=qd

→ NF (kR<1):
changes more rapidly than FF
stronger where FF vanishes

(k light wavevector, $k=\omega/c=2\pi/\lambda$)

Photon tunneling



Fresnel formulas

for refraction at the interface between 2 media

\rightarrow Near Field Optical Microscopy :

1928 : Synge's idea (lacking tech for tiny holes and scan)
1972 : first μ-wave experiments by Ash and Nichols
1984 : birth of SNOM with first visible imaging at Dieter Pohl lab in Zurich

Fiber tip formation by Heat and Pull







- + very smooth taper surface
- very low throughput (~10⁻⁷)
 due to long taper (i.e. small angle 2α~10°)

Fiber tip formation by Tube Etching



- + ease of manufacturing
- + typical etching large cone angle
- + smooth taper surface
 - compared to standard etching
- + self-limiting process
- off-center (off core?) apex
- HF vapor attacks fiber holder above



cone angle ~ 26°

NFO Microscopy setups 1/2



NFO Microscopy setups 2/2



NFO probes: aperture formation



Constant Distance Mode, but :

 NFO signal isointensity profiles not a function, e.g.:



(longitudinal, TE illumination)

 most independent distance signals have <u>off-centered tip</u>, e.g.:



detected signal

followed path

NSOM: the power of light at high resolution



contrast from:

- absorption
- reflection
- emission
- polarization

e.g. fluorescence:

- fluorochrome dye labeling, based on
 - pH or Mg²⁺ Ca²⁺
 - hydrophobic/philic interactions
 - specific bonding to prot. or nucleic acids
- Stokes shift
- τ ≤ 10 ns

+ multi-labeling - photo-bleaching

Conclusions

- a number of microscopic techniques available today:
 - conventional optical microscopy: user friendly but diffraction-limited (res.~200 nm)
 - electron microscopy: very high (~5 nm, SEM) or even atomic (TEM) resolution
 - Near Field optical microscopy:
 ~30 nm res., and spectroscopic capabilities
- our choice will depend on what we want to "see"
 - for "force" sensing, viscoelastic properties etc.:
 - \rightarrow go AFM (next seminar !)
- always be aware of artifacts:
 "the map is not the territory", Alford Korzybski

